

Weed vegetation of arable land in Central Europe: Gradients of diversity and species composition

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Abstract.

Question: What are the main broad-scale spatial and temporal gradients in species composition of arable weed communities and what are their underlying environmental variables?

Location: Czech Republic and Slovakia.

Methods: A selection of 2653 geographically stratified relevés sampled between 1954-2003 was analysed with direct and indirect ordination, regression analysis and analysis of beta diversity.

Results: Major changes in weed species composition were associated with a complex gradient of increasing altitude and precipitation and decreasing temperature and base status of the soils. The proportion of hemicryptophytes increased, therophytes and alien species decreased, species richness increased and beta diversity decreased with increasing altitude. The second most important gradient of weed species composition was associated with seasonal changes, resulting in striking differences between weed communities developed in spring and summer. In summer, weed communities tended to have more neophytes, higher species richness and higher beta diversity. The third gradient reflected long-term changes in weed vegetation over past decades. The proportion of hemicryptophytes and neophytes increased, while therophytes and archaeophytes decreased, as did species richness over time. The fourth gradient was due to crop plants. Cultures whose management involves less disturbances, such as cereals, harboured less geophytes and neophytes, and had higher species richness but lower beta diversity than frequently disturbed cultures, such as root crops.

Conclusions: Species composition of Central European weed vegetation is mainly influenced by broad-scale climatic and edaphic factors, but its variations due to seasonal dynamics and long-term changes in agricultural management are also striking. Crop plants and crop-specific management affect it to a lesser, but still significant extent.

Keywords: Alien species; Altitude; β -diversity; Canonical Correspondence Analysis; Cereal; Czech Republic; Plant community; Root crop; Seasonal dynamics; Slovakia; Temporal change.

Nomenclature: Kubát et al. (2002).

Introduction

Weed communities on arable land are widespread and highly dynamic components of Central European vegetation (Holzner 1978; Holzner & Immonen 1982; Ellenberg 1996). So far, major broad-scale patterns of weed species composition and underlying environmental gradients have been studied mainly by phytosociological methods. Earlier studies from various countries of Central and West Europe supposed the main discontinuity in weed species composition to occur between cereal and root-crop cultures, as reflected in the division of high-level phytosociological units (Braun-Blanquet et al. 1936; Tüxen 1950; Oberdorfer 1993). Studies from the last decade, which used larger data sets, supposed the main discontinuity to occur between the weed vegetation of basic soils in drier areas and of more acidic soils in precipitation-rich areas (Hüppe & Hofmeister 1990; Ries 1992; Mucina 1993; Jarolímek et al. 1997). Multivariate analyses of broad-scale gradients in similar weed communities as occur in Central Europe, carried out in northern Europe (Salonen 1993; Erviö et al. 1994; Andersson & Milberg 1998; Hallgren et al. 1999) or southern Canada (Thomas & Dale 1991; Dale et al. 1992) also emphasized the role of geographical variability in climate and soil, which is usually more important for weed species composition than the effect of crops and associated agricultural management. For Central European weed vegetation, however, such an analysis has never been done.

Weed vegetation, consisting predominantly of annual plants, shows a much higher degree of temporal dynamics than other vegetation types in Central Europe. These dynamics operate both on the scale of seasonal changes (Kropáč et al. 1971; Holzner 1973, 1978; Lososová et al. 2003) and on a long-term scale, that corresponds to the gradually increasing intensification of agricultural production during the second half of the 20th century (Hilbig 1987; Kropáč 1988; Andreasen et al. 1996). Thus, gradients in

species composition of weed vegetation cannot be properly investigated without consideration of temporal patterns.

This paper applies multivariate statistical methods to a large data set of weed relevés from two Central European countries. Our objectives were to test the effects of broad-scale environmental gradients, crop plants and time on weed species composition, to rank the importance of these particular factors, and to describe patterns in species composition and diversity of weed vegetation associated with these factors.

Materials

We compiled a data set of 3481 relevés of agricultural weed vegetation of the Czech Republic (2596 relevés) and Slovakia (885 relevés). This data set included 1826 unpublished relevés of our own (in particular by Z.K., Z.O., Š.C. and Z.L.) and relevés taken from literature (e.g. Passarge & Jurko 1975; Mochňáký 1987; Otýpková 2001;) and other sources. The relevés are currently stored in TURBOVEG format (Hennekens & Schaminée 2001) in the Czech and Slovak national phytosociological databases (Chytrý & Rafajová 2003). For the analysis, we deleted all relevés lacking any indication of locality, date of record and crop plant. To achieve a reasonable stratification of the data set by geographical areas and by different vegetation types, we divided the area of the Czech Republic and Slovakia into grid squares of 1.25 longitudinal \times 0.75 latitudinal minute (ca. 1.5 km \times 1.4 km) and we also divided the relevés into spring (recorded before 1 June) and summer/autumn subsets (recorded on 1 June or later). If two or more relevés were recorded in the same grid square, the same crop plant and in the same period (spring or summer/autumn), we randomly selected only one of them. This procedure eliminated possible bias in the data set that may have been caused by oversampling of some areas, and yielded 2653 relevés, which were further used in analyses. These relevés were recorded between 1954–2003, mostly in the periods 1960–1985 and 1995–2003. Plot sizes ranged from 8 to 100 m².

Records of crop plants were deleted from the species data set and used as explanatory variables. A few plant records determined only at the genus level were deleted. Bryophytes were not considered as they play a negligible role in agricultural habitats and often are not recorded by researchers. Species cover values, estimated at different ordinal scales in original relevés, were transformed to percentages and square-rooted.

Four explanatory variables, presumably associated with variation in weed species composition, were compiled for all relevés: Altitude, Season, Year of record and Crop. The variable Season expressed phenological stages

of weed vegetation during the growing season. It was derived from the date of record transformed into sequential numbers of half-month periods from the beginning of the year. For instance, 9 and 14 were the values used for the first half of May and the second half of July, respectively. Variable Crop included two broad categories strongly differing in agricultural management, further referred to as Cereals and Root crops, the former without mechanical disturbances applied during the period of crop growth while the latter were subject to regular hoeing, weeding or tilling. 'Cereals' included cereals itself (1355 relevés), recently abandoned fields (206), fodder (128), maize (119), stubble (92), and rapeseed (69) and the category 'Root crops' involved root crops themselves (427), vegetables (95), vineyards (86), and other crops (76).

Methods

First, we subjected the weed data set to Detrended Correspondence Analysis (DCA) from the CANOCO 4.5 package (ter Braak & Šmilauer 2002), in order to assess the overall variation patterns in species composition. DCA, a unimodal method, was used due to rather long gradients in compositional turnover encountered within the data set. For interpretation of DCA results in terms of environmental gradients, the four explanatory variables (Altitude, Season, Year, and Crop) were passively projected on an ordination scatter plot. These variables, however, were not involved in the extraction of ordination axes and the axes therefore correspond to the main gradients in species composition, that are unconstrained by explanatory variables.

Subsequently we tested gross and net effects of each of the four explanatory variables on species composition using canonical correspondence analysis (CCA; ter Braak & Šmilauer 2002). Gross effects were tested using separate CCAs with a single explanatory variable, followed by permutation tests for the first canonical axis (999 permutations were always used). Net effects, i.e. the effects of particular variables after partialling out the effects shared with the other explanatory variables, were tested using partial CCAs, each with a single explanatory variable and the other three variables used as covariables. Significances were again tested by permutation tests for the first canonical axis. Shared effects of different combinations of explanatory variables were tested using a series of partial CCAs. The ratio of particular canonical eigenvalues to the sum of all eigenvalues (total inertia) was used to measure the proportion of explained variation (Borcard et al. 1992). For the four partial CCAs in which a single explanatory variable was used, we listed scores along the first CCA axis for species with the highest fit in the analysis. The resulting species order reflected the

weed vegetation change along the gradient of the particular explanatory variable, after partialling out effects of the other variables. Other possible gradients in species composition that could not be explained by the four variables were assessed by partial DCA, in which these four variables were used as covariables.

Ecological interpretations of the gradients associated with the net effects of each explanatory variable were done by relating these gradients to Ellenberg indicator values for relevés (Ellenberg et al. 1992). To investigate structural changes in vegetation along these gradients, in addition to species composition, we used species richness and species classifications by life forms and by time of immigration for alien species. As the plot size varied in our data set, number of species was replaced by residuals from the regression of the number of species on log plot size, in order to remove variance in species numbers due to species-area relationship (Rosenzweig 1995). In the subsequent analyses, these residuals were used as a measure of species richness. Although the area effect was removed, the results for species richness must be interpreted with caution due to the non-random arrangement of sampling sites (Chytrý 2001). However, as we found no systematic bias in plot size with respect to environmental gradients, we believe that the patterns of species richness detected in our data set approximate the real situation.

Raunkiaer life forms were taken mostly from Ellenberg et al. (1992). Only therophytes, hemicryptophytes and geophytes were used in the analyses, as the other life forms were rare in weed vegetation. Alien species were classified according to Pyšek et al. (2002a) into archaeophytes (non-native species that appeared in the study area before 1500, and many of them already with the advent of Neolithic agriculture) and neophytes (non-native species that appeared during the last five centuries).

The net effects of each of the explanatory variables Altitude, Season, Year and Crop (the effects after subtracting correlated effects of the other three variables) were calculated for species richness and Ellenberg indicator values as dependent variables. For quantitative variables Altitude, Season and Year, net effects were determined using standardized residuals from general linear models in which each of these three variables was in turn used as a dependent variable and the other two plus categorical variable Crop as independent variables. Then species richness and Ellenberg indicator values were regressed on these standardized residuals. Standardization of the residuals enabled direct comparison of regression coefficients among explanatory variables measured on different scales. Net effects of categorical variable Crop were determined using analysis of covariance.

Relationships between life forms or groups of alien species and the net effects of the four explanatory variables were tested with a generalized linear model, using binomial

distribution and logit link function (McCullagh & Nelder 1989), where the independent variable was species scores on the 1st canonical axis of partial CCA with the given explanatory variable and dependent categorical variable was species assignment to a particular category of life form or alien status. Only species whose fit in the partial CCA was higher than median were used in the analysis. All univariate statistical analyses in this study were calculated with the STATISTICA program (Anon. 2001).

In order to assess patterns of β -diversity (the mean difference in weed species composition among relevés) along the four major gradients, we partitioned the data set along the gradients, and for each partition we calculated β -diversity as mean Sørensen dissimilarity for all pairs of relevés ($1 - S$, where S is Sørensen similarity; Magurran 1988; Koleff et al. 2003). Then we determined confidence intervals for β -diversity in each partition, using 500 bootstrap samples (Efron & Tibshirani 1993) taken from the relevés belonging to that partition. This procedure was done in program JUICE 6.1 (Tichý 2002).

Results

Fig. 1 shows the variation in weed species composition, as detected by DCA. The first ordination axis explained 1.73% of the total variation in species data and corresponded to the altitudinal gradient, while the second axis (1.46% of total variation) was associated with other explanatory variables. The amount of variation in species data explained by net effects of particular variables, as detected by partial CCAs (Table 1), was highest for Altitude and decreased through Season and Year to Crop. The explained variation attributable to net shared effects of two or three variables was negligible. These four variables together explained 2.30% of the total variation in species data. The first and the second axis of partial

Table 1. Percentage variation in species data attributed to the effects of explanatory variables, as calculated by (partial) CCA. Gross effects include the total variation explained by the particular variable(s), including the variation shared with other variables. Net effects include variation explained by the particular variable(s), not shared with other variables. Percentage of net shared variation (\cap) is only shown when exceeding 0.01%. *F*-values of the permutation tests are shown for partial CCAs, which were used to determine the net effects. ** = $P < 0.01$.

	Gross effects	Net effects	Permutation test: <i>F</i> -value
Altitude	0.84	0.84	22.760 **
Season	0.69	0.61	16.546 **
Year	0.51	0.48	13.201 **
Crop	0.37	0.26	7.143 **
Season \cap Crop	-	0.08	
Year \cap Crop	-	0.03	
All explanatory variables	2.30	2.30	

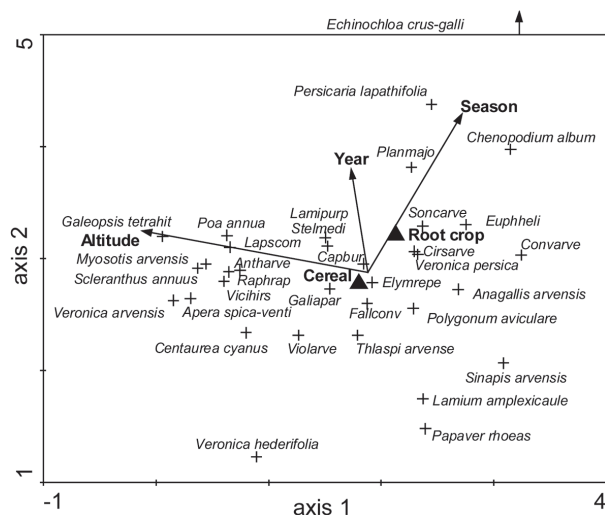


Fig. 1. Detrended correspondence analysis (DCA) diagram of species with passively projected explanatory variables for weed vegetation of the Czech Republic and Slovakia. Species with low weight are not shown. Abbreviations: Antharve = *Anthemis arvensis*; Capbur = *Capsella bursa-pastoris*; Cirsarve = *Cirsium arvense*; Convarve = *Convolvulus arvensis*; Elymrepe = *Elytrigia repens*; Eupheli = *Euphorbia helioscopia*; Fallconv = *Fallopia convolvulus*; Galiapar = *Galium aparine*; Lamipurp = *Lamium purpureum*; Lapscomm = *Lapsana communis*; Planmajo = *Plantago major*; Raphrap = *Raphanus raphanistrum*; Soncave = *Sonchus arvensis*; Stelmedi = *Stellaria media*; Vicihirs = *Vicia hirsuta*; Violarve = *Viola arvensis*.

DCA, calculated after partialling out the variation explained by the four explanatory variables, explained additional 1.59% and 1.10% of the total variation, respectively. Comparable proportions of the total variation explained by variables Altitude, Season, Year and Crop and by partial DCA indicate that these four variables are among the most important variables which affect weed species composition, however, there may also be a few other explanatory variables of comparable importance.

In spite of the low percentage of explained variation, the effects of each of these variables on species composition were highly significant and species ranks along the gradients of each variable had clear ecological interpretations (Table 2; next page). Interpretations of these gradients through Ellenberg indicator values are presented in Table 3. Beside changes in species composition, these four gradients were also correlated with other vegetation characteristics. Species richness was higher at higher altitudes, later in the growing season and in cereal fields, but decreased over the years. β -diversity (Fig. 2) decreased with altitude but increased from spring to summer and autumn. It was higher in root crops than in cereals, but showed no interpretable pattern of change over the years. Therophytes were more common at lower altitudes and in earlier decades of the second half of the

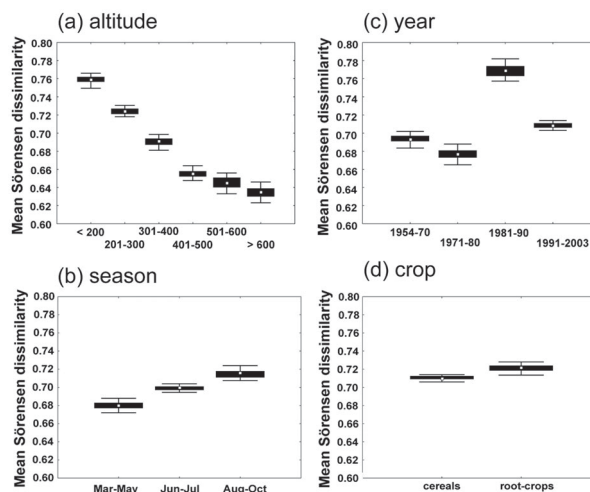


Fig. 2. Differences in β -diversity along four major gradients. β -diversity increases with mean pair-wise Sørensen dissimilarity calculated for groups of relevés. Boxes and whiskers indicate median and 50% and 95% percentiles, calculated from bootstrap resampling.

20th century, while hemicryptophytes showed opposite patterns. Geophytes appeared to be more frequent in root crops (Table 3). Representation of alien species, both archaeophytes and neophytes, decreased with altitude (Table 3). Archaeophytes exhibited a remarkable decline over the years, while neophytes tended to be progressively more common. Neophytes were especially found in root-crops and later in the growing season.

Table 3. Relationships between net effects of the four explanatory variables and vegetation characteristics. Relationships with Ellenberg indicator values and diversity measures were calculated by partial correlation coefficients. Relationships with life forms and alien species were expressed as coefficients of the generalized regression model. In categorical variable Crop, signs of inequality indicate significantly higher values found in either cereals (C > R) or root crops (C < R). ** = $P < 0.01$; * = $P < 0.05$; n.s. = non significant.

	Altitude	Season	Year	Crop
Ellenberg indicator values				
Light	-0.056**	0.043**	0.010 *	C < R**
Temperature	-0.111**	0.032**	-0.012**	C < R**
Continentality	-0.155**	0.020 *	-0.044**	n.s.
Moisture	0.132**	0.040**	0.093**	n.s.
Soil reaction	-0.284**	-0.031 *	0.083**	n.s.
Nutrients	-0.067**	0.036**	0.229**	C < R**
Species richness	1.376**	2.384**	-0.768**	C > R**
Life forms				
Therophytes	-0.777**	n.s.	-1.126**	n.s.
Hemicryptophytes	0.677**	n.s.	1.074**	n.s.
Geophytes	n.s.	n.s.	n.s.	C < R*
Alien species				
Archaeophytes	-0.797**	n.s.	-0.641**	n.s.
Neophytes	-0.715**	0.575 *	0.732 *	C < R**

Discussion

Low proportion of explained variation

The present analysis identified main gradients in weed species composition and interpreted them in terms of explanatory variables. Although the gradients had clear ecological interpretations, the variation explained by individual ordination axes was extremely low (< 2%). This could be partly attributed to the fact that some important explanatory variables, such as the effects of herbicides, fertilizing, crop rotation or land use in preceding years, were not considered in our analysis. In search for more powerful explanatory variables, we divided the Crop variable into more categories, but the amount of explained variation did not increase significantly and the results had a much less straightforward interpretation, therefore they are not presented here. Beside the low amount of total variation explained by variables Altitude, Season, Year and Crop, it was striking that unconstrained and partial unconstrained ordination (DCA, pDCA) also detected low variation, which suggested that the potentially missing variables were hardly more important than the four variables included. Thus the low proportion of explained variation was perhaps a consequence of (1) the large data set (2653 relevés × 544 species), resulting in a high amount of noise, and (2) the polynomial distortion of ordination axes, which had been shown by Økland (1999) to increase the total inertia and thus to underestimate the proportion of explained variation. Therefore we use the eigenvalue/total inertia ratio for comparison of the relative importance of individual explanatory variables or ordination axes rather than as real proportion of explained variation.

The effect of altitude

The most important gradient in species composition of weed vegetation covaried with altitude and associated climatic factors (temperature, precipitation) (Table 3). In the Czech Republic and less so in Slovakia the pH and nutrient status of soils is also correlated with altitude, as base-rich soils are mainly found in dry lowlands. Therefore there is a clear distinction between thermophilous, xerophilous and calcicole weed communities at low altitudes and communities of colder and wetter areas with acidic soils at higher altitudes. This result demonstrates that at the broad geographical scale, even the vegetation of a human-made habitat containing a large proportion of alien species and strongly depending on management, is more influenced by primary environmental factors than human activities. Similar results, which stressed the importance of climatic or soil variables on weed vegetation,

Table 2. Species with the highest fit in partial CCAs and their scores along ordination axis 1. Each column represents a separate partial CCA in which Altitude, Season, Year, and Crop were used in turn as the only explanatory variable while the effects of the other variables were subtracted by defining them as covariables. Low values of scores for respective variables are associated in turn with low altitudes, early season, earlier years and cereal fields. For an extended version of this table, see App. 1. *Veronica hederifolia* includes *V. sublobata*.

Altitude Species	Season Species	Year Species	Crop Species
<i>Setaria pumila</i>	<i>Veronica triphyllus</i>	<i>Myosotis stricta</i>	<i>Apera spica-venti</i>
<i>Amaranthus retroflexus</i>	<i>Erophila verna</i>	<i>Silene latifolia</i>	<i>Euphorbia exigua</i>
<i>Echinochloa crus-galli</i>	<i>Veronica hederifolia</i> ¹	<i>Papaver argemone</i>	<i>Consolida regalis</i>
<i>Consolida regalis</i>	<i>Myosotis stricta</i>	<i>Neslia paniculata</i>	<i>Silene noctiflora</i>
<i>Veronica polita</i>	<i>Papaver argemone</i>	<i>Arenaria verpyllifolia</i>	<i>Vicia tetrasperma</i>
<i>Silene noctiflora</i>	<i>Arabisopsis thaliana</i>	<i>Raphanus raphanistrum</i>	<i>Vicia sativa</i>
<i>Myosotis arvensis</i>	<i>Lamium amplexicaule</i>	<i>Euphorbia exigua</i>	<i>Anagallis arvensis</i>
<i>Veronica arvensis</i>	<i>Descurainia sophia</i>	<i>Sherardia arvensis</i>	<i>Myosotis arvensis</i>
<i>Raphanus raphanistrum</i>	<i>Lamium purpureum</i>	<i>Scleranthus annuus</i>	<i>Polygonum aviculare</i>
<i>Vicia sativa</i>	<i>Thlaspi arvense</i>	<i>Rumex crispus</i>	<i>Viola arvensis</i>
<i>Achillea millefolium</i>	<i>Stellaria media</i>	<i>Taraxacum sect. Ruderalia</i>	<i>Euphorbia helioscopia</i>
<i>Lapsana communis</i>	<i>Capsella bursa-pastoris</i>	<i>Gallium aparine</i>	<i>Stellaria media</i>
<i>Ranunculus repens</i>	<i>Gallium aparine</i>	<i>Lapsana communis</i>	<i>Chenopodium album</i>
<i>Mentha arvensis</i>	<i>Anagallis arvensis</i>	<i>Ranunculus repens</i>	<i>Persicaria lapathifolia</i>
<i>Tussilago farfara</i>	<i>Plantago major</i>	<i>Lamium purpureum</i>	<i>Echinochloa crus-galli</i>
<i>Spergula arvensis</i>	<i>Medicago lupulina</i>	<i>Lamium dissectum</i>	<i>Amaranthus retroflexus</i>
<i>Rumex acetosella</i>	<i>Sonchus oleraceus</i>	<i>Poa trivialis</i>	<i>Gallinsoga parviflora</i>
<i>Galeopsis tetrahit</i>	<i>Sonchus asper</i>	<i>Aegopodium podagraria</i>	<i>Solanum nigrum</i>
<i>Holcus mollis</i>	<i>Euphorbia exigua</i>	<i>Lolium perenne</i>	<i>Gallinsoga quadriradiata</i>
<i>Alopecurus vulgaris</i>	<i>Setaria pumila</i>	<i>Gallinsoga quadriradiata</i>	<i>Senecio vulgaris</i>
Ax 1 score	Ax 1 score	Ax 1 score	Ax 1 score
-0.97	-0.97	-1.41	-0.87
-0.94	-0.94	-1.35	-0.78
-0.94	-0.94	-1.24	-0.76
-0.63	-0.63	-1.07	-0.60
-0.62	-0.62	-0.84	-0.54
-0.49	-0.49	-0.73	-0.53
0.33	0.33	-0.70	-0.43
0.39	0.39	-0.70	-0.42
0.41	0.41	-0.46	-0.41
0.42	0.42	-0.38	-0.37
0.51	0.51	-0.29	-0.28
0.52	0.52	-0.27	0.28
0.53	0.53	-0.25	0.29
0.55	0.55	0.36	0.32
0.71	0.71	0.38	0.34
0.79	0.79	0.40	0.43
0.81	0.81	0.48	0.53
0.91	0.91	0.53	0.65
1.32	1.32	0.54	0.65
2.12	2.12	0.87	1.06
Fit	Fit	Fit	Fit
0.057	0.057	0.072	0.029
0.058	0.058	0.053	0.039
0.072	0.072	0.201	0.028
0.072	0.072	0.044	0.061
0.065	0.065	0.035	0.039
0.060	0.060	0.065	0.086
0.089	0.089	0.118	0.029
0.051	0.051	0.043	0.029
0.052	0.052	0.068	0.037
0.063	0.063	0.089	0.038
0.080	0.080	0.061	0.045
0.116	0.116	0.067	0.051
0.074	0.074	0.038	0.044
0.075	0.075	0.086	0.044
0.053	0.053	0.36	0.031
0.063	0.063	0.38	0.058
0.094	0.094	0.40	0.033
0.063	0.063	0.48	0.037
0.326	0.326	0.53	0.031
0.086	0.086	0.46	0.062
0.063	0.063	0.87	0.073
Fit	Fit	Fit	Fit
0.025	0.025	0.072	0.029
0.022	0.022	0.053	0.039
0.024	0.024	0.201	0.028
0.020	0.020	0.044	0.061
0.015	0.015	0.035	0.039
0.020	0.020	0.065	0.086
0.020	0.020	0.118	0.029
0.023	0.023	0.043	0.029
0.022	0.022	0.068	0.037
0.022	0.022	0.089	0.038
0.014	0.014	0.061	0.045
0.029	0.029	0.067	0.051
0.049	0.049	0.038	0.044
0.023	0.023	0.36	0.031
0.021	0.021	0.38	0.058
0.023	0.023	0.40	0.033
0.030	0.030	0.48	0.037
0.016	0.016	0.53	0.031
0.031	0.031	0.54	0.062
0.035	0.035	0.87	0.073

were reported by studies from other geographical areas, such as Manitoba and Saskatchewan (Thomas & Dale 1991; Dale et al. 1992), Finland (Salonen 1993; Erviö et al. 1994) and Sweden (Hallgren et al. 1999). The dominant role of climate and soil in forming broad-scale compositional patterns of weed vegetation in Central Europe was discussed by Holzner (1978) and recognized in recent phytosociological surveys (Hüppe & Hofmeister 1990; Ries 1992; Mucina 1993; Jarolímek et al. 1997).

Altitude also influences the diversity of weed vegetation. Unexpectedly, species richness increases with altitude, which is in contrast with the decreasing patterns that are more commonly found in temperate regions (Begon et al. 1990; Pyšek et al. 2002c). Higher numbers of species per relevé plot at higher altitudes are probably due to a lower degree of agricultural intensification in less productive upland areas. By contrast, β -diversity, i.e. between-site variation in species composition, decreases with altitude (Fig. 2a). This pattern is consistent with the contrast observed between the remarkable seasonal dynamics and pronounced differentiation due to crop plants in the lowlands and warmer parts of Europe on the one hand and the rather uniform floristic composition throughout the season and across different crop plants in the highlands and cooler parts of Europe on the other hand (Holzner 1978; Holzner & Immonen 1982; Glemnitz et al. 2000). Similar relationships of floral and vegetation diversity to altitude as found in this study were reported for Central European urban vegetation (Pyšek 1993).

Altitude and associated environmental factors do not only influence species composition and diversity, but also a proportion of dominant life forms of weed vegetation, therophytes and hemicryptophytes. The proportional change from therophytes to hemicryptophytes along the altitudinal gradient, revealed in our study, suggests that at higher altitudes weed communities contain more species of adjacent vegetation, e.g. meadows and pastures, while the performance of ecologically specialized annual weeds decreases.

Our study also demonstrated that alien weeds, both archaeophytes and neophytes, showed a remarkable concentration of occurrences at lower altitudes, which corresponds to analogous patterns found in various vegetation types of Central Europe (Kowarik 1990; Mihulka 1998; Pyšek 1998a, b, 2002b; Sukopp 2002). In the context of the present paper this is not surprising for neophytes, recent newcomers from regions with often a warmer climate than the target area of Central Europe (Pyšek et al. 2003). However, our results indicate that also for archaeophytes, a group with thousands of years of invasion history in the territory studied, climate still acts as a major constraint to their wider distribution even in habitats to which they are perfectly suited.

Seasonal changes

The second most important gradient in weed species composition is related to seasonal dynamics. Short-living annual weeds form remarkably different communities in the course of a single growing season, with late-germinating thermophilous species appearing only in late spring and summer (Ellenberg 1996). In a phytosociological context, this observation led Kropáč et al. (1971) and Holzner (1973, 1978) to a proposal for a classification of spring, summer and autumn phenological aspects found on the same field as separate associations. The dynamics of thermophilous annuals, many of them neophytes, probably cause the increase in species richness and β -diversity during the growing season (Table 3, Fig. 2b). Increasing performance of heliophilous plants later in the season (Table 3) is probably caused by an increased light availability after crop harvest.

Changes over past decades

The third gradient in species composition is a historical gradient, reflecting the changes of weed vegetation over past decades. Our data set contained many relevés from the 1960s, as well as many relevés made after the mid-1990s, but only 13 relevés were from before 1960. This means that the vast majority of our relevés were from the period when most agricultural land in former Czechoslovakia was owned by large co-operative farms that had changed small fields into vast tracts of arable land with intensive management and heavy use of herbicides. This management resulted in a dramatic decline of diversity of Central European weed vegetation (Kropáč 1988; Hilbig & Bachthaler 1992; Lososová 2003). It is therefore possible that temporal trends in weed vegetation were not so striking as they would have been if relevés from the 1940s would have been involved in the comparison. In this respect, our results were similar to those of Hallgren et al. (1999), who found weak temporal trends in Swedish weed data from 1970-1994. Still, species order along the time axis (Table 2) and regression analyses (Table 3) indicate a decline in archaeophytic annuals (e.g. *Papaver argemone*, *Neslia paniculata*, *Raphanus raphanistrum*, *Euphorbia exigua*, *Sherardia arvensis*) and increase in neophytes, which is most striking in the American species *Galinsoga quadriradiata*. The overall decrease in species richness over time (Table 3) confirmed the results of the above mentioned studies on biodiversity decline in Central European weed vegetation. The pattern of β -diversity through time (Fig. 2c) had no clear interpretation. A pronounced peak in the 1980s was probably influenced by a lower intensity of sampling and associated bias in this period.

The effect of the crop plant

The type of crop affected weed species composition: thermophilous and nutrient-demanding weeds were more frequent in root crops. Cereal fields had higher species richness, lower β -diversity (Fig. 2d), and fewer geophytes and neophytes than root crop cultures (Table 3). The effect of the crop plant, however, was less pronounced than the effects of altitude, season or year (Table 1). This result contradicts the earlier phytosociological concepts of two major units of Central European weed vegetation, one for cereal fields and the other for root crops. Such a distinction has been made either at the level of classes, e.g. *Secalietea* and *Chenopodietaea*, or orders within a single class, mostly the class *Stellarietea mediae* (e.g. Braun-Blanquet et al. 1936; Tüxen 1950; Mucina & Maglocký 1985; Krippelová & Mucina 1988; Kropáč 1988; Oberdorfer 1993; Moravec et al. 1995). This contradiction indicates that in the traditional phytosociological approach, emphasis on a subjectively selected environmental factor such as the crop plant may lead to a failure to identify properly the main gradients in species composition, especially in a vegetation type driven simultaneously by several environmental factors. Statistical analyses of large data sets appear to be a more suitable tool for identifying the importance of particular environmental factors.

The overriding effect of climatic and soil factors revealed in the current study is in accordance with the results of Hüppe & Hofmeister (1990) who proposed a new syntaxonomical system of German weed vegetation with two main groups of communities, i.e. calcifuge and calcicole. Similar divisions, which put less emphasis on the differentiation of weed vegetation due to the crop, were accepted by Ries (1992) and Mucina (1993) for Austria, Jarolímek et al. (1997) for Slovakia, and Haveman et al. (1998) for The Netherlands.

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App. 1. Extended version of Table 2. Species with the highest fit in partial CCAs and their scores along the first ordination axes. Each column represents a separate partial CCA in which Altitude, Season, Year, and Crop were used in turn as the only explanatory variable while the effects of the other variables were subtracted by defining them as covariables. Low values of scores for respective variables are associated in turn with low altitudes, early season, earlier years and cereal fields.

Altitude Species	Ax 1 score	Fit	Season Species	Ax 1 score	Fit	Year Species	Ax 1 score	Fit	Crop Species	Ax 1 score	Fit
<i>Portulaca oleracea</i>	-1.61	0.010	<i>Cerastium brachypetalum</i>	-2.43	0.007	<i>Agrostemma githago</i>	-1.15	0.014	<i>Galeopsis angustifolia</i>	-0.79	0.003
<i>Diphloxia muralis</i>	-1.05	0.009	<i>Tragopogon dubius</i>	-2.03	0.005	<i>Galium tricornutum</i>	-1.14	0.017	<i>Trifolium medium</i>	-0.55	0.003
<i>Senecio pinnatifidus</i>	-0.96	0.057	<i>Gagea pratensis</i>	-1.60	0.006	<i>Ranunculus scordium</i>	-0.93	0.007	<i>Kickxia elatiae</i>	-0.64	0.014
<i>Amaranthus retroflexus</i>	-0.94	0.058	<i>Halostemum unbelatum</i>	-1.53	0.021	<i>Galeopsis ladanum</i>	-0.92	0.013	<i>Cerastium glomeratum</i>	-0.55	0.003
<i>Echinochloa crus-galli</i>	-0.94	0.072	<i>Veronica triphyllos</i>	-1.46	0.034	<i>Plantago media</i>	-0.91	0.016	<i>Agrostis gigantea</i>	-0.54	0.005
<i>Corys canadensis</i>	-0.93	0.038	<i>Veronica triphyllos</i>	-1.41	0.072	<i>Crepis capillaris</i>	-0.91	0.010	<i>Mispopates orontium</i>	-0.53	0.008
<i>Mercurialis annua</i>	-0.91	0.016	<i>Erophila verna</i>	-1.35	0.053	<i>Mysotis discolor</i>	-0.89	0.009	<i>Galeopsis ladanum</i>	-0.51	0.004
<i>Solanum nigrum</i>	-0.89	0.026	<i>Veronica praecox</i>	-1.32	0.014	<i>Mysotis stricta</i>	-0.87	0.029	<i>Tanacetum vulgare</i>	-0.45	0.003
<i>Ajuga reptans</i>	-0.88	0.011	<i>Veronica hederifolia</i>	-1.24	0.201	<i>Rubus fruticosus</i> agg.	-0.85	0.018	<i>Dianthus barbatus</i>	-0.42	0.007
<i>Amaranthus powellii</i>	-0.86	0.010	<i>Thlaspi perfoliatum</i>	-1.23	0.024	<i>Cornifolia orientalis</i>	-0.84	0.014	<i>Apera spica-venti</i>	-0.41	0.025
<i>Gypsophila muralis</i>	-0.85	0.017	<i>Bromus tectorum</i>	-1.10	0.005	<i>Misopates orontium</i>	-0.78	0.017	<i>Phleum pratense</i>	-0.41	0.006
<i>Euphorbia falcata</i>	-0.84	0.021	<i>Mysotis stricta</i>	-1.07	0.044	<i>Stene latifolia</i>	-0.78	0.039	<i>Lolium multiflorum</i>	-0.40	0.003
<i>Reseda laterola</i>	-0.81	0.011	<i>Mysotis minimus</i>	-1.03	0.008	<i>Alchemilla vulgaris</i>	-0.76	0.008	<i>Juncus bufonius</i>	-0.39	0.005
<i>Stachys foetida</i>	-0.80	0.026	<i>Bromus sterilis</i>	-0.89	0.007	<i>Papaver argemone</i>	-0.76	0.028	<i>Euphorbia falcata</i>	-0.38	0.004
<i>Stachys annua</i>	-0.78	0.032	<i>Papaver argemone</i>	-0.84	0.035	<i>Veronica triphyllos</i>	-0.75	0.020	<i>Trifolium arvense</i>	-0.38	0.006
<i>Cardaria draba</i>	-0.78	0.016	<i>Valerianella locusta</i>	-0.78	0.009	<i>Erophila verna</i>	-0.72	0.015	<i>Caucalis platycarpus</i>	-0.38	0.004
<i>Senecio viridiflorus</i>	-0.77	0.031	<i>Mysotis discolor</i>	-0.77	0.006	<i>Veronica triloba</i>	-0.71	0.008	<i>Euphorbia exigua</i>	-0.37	0.022
<i>Kickxia spuria</i>	-0.74	0.019	<i>Arabisopsis thaliana</i>	-0.73	0.065	<i>Gypsophila muralis</i>	-0.67	0.011	<i>Trifolium campestre</i>	-0.37	0.006
<i>Cheopodium hybridum</i>	-0.74	0.024	<i>Lithospermum arvense</i>	-0.70	0.031	<i>Papaver dubium</i>	-0.64	0.012	<i>Consolida regalis</i>	-0.36	0.024
<i>Galiniasa parviflora</i>	-0.72	0.032	<i>Laminium amplexicaule</i>	-0.70	0.118	<i>Veronica opaca</i>	-0.63	0.007	<i>Aphanes arvensis</i>	-0.36	0.012
<i>Arctium tomentosum</i>	-0.71	0.010	<i>Discarumia sophia</i>	-0.70	0.043	<i>Sagina procumbens</i>	-0.63	0.014	<i>Ranunculus arvensis</i>	-0.31	0.005
<i>Descurainia sophia</i>	-0.71	0.045	<i>Adonis aestivialis</i>	-0.63	0.018	<i>Spergularia rubra</i>	-0.62	0.007	<i>Adonis aestivialis</i>	-0.29	0.004
<i>Cichorium intybus</i>	-0.68	0.010	<i>Anthemis austriaca</i>	-0.59	0.010	<i>Linaria vulgaris</i>	-0.60	0.025	<i>Stene noctiflora</i>	-0.28	0.020
<i>Anthemis austriaca</i>	-0.65	0.012	<i>Fumaria vallonit</i>	-0.54	0.009	<i>Neslia paniculata</i>	-0.60	0.061	<i>Matricaria recutita</i>	-0.28	0.004
<i>Lactuca serriola</i>	-0.64	0.024	<i>Cornifolia orientalis</i>	-0.53	0.005	<i>Trifolium arvense</i>	-0.60	0.015	<i>Sagina procumbens</i>	-0.28	0.003
<i>Consolida regalis</i>	-0.63	0.072	<i>Cardaria draba</i>	-0.49	0.006	<i>Vicia villosa</i>	-0.58	0.010	<i>Vicia tetrasperma</i>	-0.28	0.015
<i>Veronica poliflora</i>	-0.62	0.065	<i>Laminium purpureum</i>	-0.46	0.068	<i>Hypolepidium maximum</i>	-0.57	0.008	<i>Poa trivialis</i>	-0.28	0.007
<i>Falcaria vulgaris</i>	-0.57	0.009	<i>Papaver dubium</i>	-0.43	0.005	<i>Adonis aestivialis</i>	-0.55	0.013	<i>Stachys annua</i>	-0.28	0.004
<i>Euphorbia exigua</i>	-0.55	0.049	<i>Veronica poliflora</i>	-0.40	0.027	<i>Arenaria serpyllifolia</i>	-0.54	0.039	<i>Centaura cyanus</i>	-0.26	0.012
<i>Microthymum thymus</i>	-0.54	0.017	<i>Thlaspi arvense</i>	-0.38	0.089	<i>Raphanus raphanistrum</i>	-0.53	0.086	<i>Galium spurium</i>	-0.25	0.004
<i>Adonis aestivialis</i>	-0.51	0.012	<i>Ranunculus arvensis</i>	-0.38	0.007	<i>Cardaria draba</i>	-0.51	0.007	<i>Aethusa cynapium</i>	-0.24	0.012
<i>Stene noctiflora</i>	-0.49	0.060	<i>Poa trivialis</i>	-0.38	0.012	<i>Anthemis austriaca</i>	-0.51	0.008	<i>Papaver rhoas</i>	-0.24	0.014
<i>Lathyrus tuberosus</i>	-0.47	0.031	<i>Papaver rhoas</i>	-0.37	0.031	<i>Trifolium campstre</i>	-0.49	0.011	<i>Vicia sativa</i>	-0.24	0.004
<i>Papaver rhoas</i>	-0.45	0.047	<i>Consolida regalis</i>	-0.34	0.021	<i>Valerianella dentata</i>	-0.49	0.025	<i>Vicia sativa</i>	-0.23	0.020
<i>Laminium amplexicaule</i>	-0.39	0.036	<i>Lolium perenne</i>	-0.32	0.006	<i>Lithospermum arvense</i>	-0.45	0.013	<i>Valerianella dentata</i>	-0.22	0.005
<i>Arenaria serpyllifolia</i>	-0.34	0.010	<i>Neslia paniculata</i>	-0.31	0.017	<i>Euphorbia exigua</i>	-0.43	0.029	<i>Daucus carota</i>	-0.20	0.007
<i>Sonchus oleraceus</i>	-0.33	0.019	<i>Fumaria officinalis</i>	-0.31	0.061	<i>Erodium cicutarium</i>	-0.42	0.025	<i>Medicago lupulina</i>	-0.19	0.008
<i>Cheopodium polyspermum</i>	-0.33	0.012	<i>Stellaria media</i>	-0.29	0.062	<i>Sierardia arvensis</i>	-0.42	0.029	<i>Veronica arvensis</i>	-0.19	0.012
<i>Sinapis arvensis</i>	-0.29	0.024	<i>Capsetta bursa-pastoris</i>	-0.27	0.067	<i>Scleranthus annuus</i>	-0.41	0.037	<i>Descurainia sophia</i>	-0.19	0.005
<i>Sonchus asper</i>	-0.29	0.027	<i>Aphanes arvensis</i>	-0.25	0.006	<i>Arabisopsis thaliana</i>	-0.39	0.019	<i>Lathyrus tuberosus</i>	-0.18	0.003
<i>Arenaria serpyllifolia</i>	-0.29	0.011	<i>Galium aparine</i>	-0.25	0.038	<i>Consolida regalis</i>	-0.39	0.027	<i>Anagallis arvensis</i>	-0.17	0.020
<i>Medicago lupulina</i>	-0.28	0.044	<i>Geranium pusillum</i>	-0.24	0.010	<i>Rumex crispus</i>	-0.37	0.038	<i>Lapsana communis</i>	-0.17	0.012
<i>Cheopodium album</i>	-0.24	0.039	<i>Poa annua</i>	-0.21	0.013	<i>Aphanes arvensis</i>	-0.35	0.011	<i>Vicia hirsuta</i>	-0.17	0.008
<i>Anagallis arvensis</i>	-0.19	0.024	<i>Veronica arvensis</i>	-0.21	0.015	<i>Gnaphalium uliginosum</i>	-0.34	0.014	<i>Matricaria discoidata</i>	-0.17	0.003
<i>Fallopia convolvulus</i>	0.10	0.013	<i>Viola arvensis</i>	-0.15	0.030	<i>Laminium amplexicaule</i>	-0.32	0.025	<i>Mysotis arvensis</i>	-0.17	0.023
<i>Viola arvensis</i>	0.12	0.020	<i>Taraxacum sect. Ruderalia</i>	-0.11	0.007	<i>Rumex acetosella</i>	-0.31	0.010	<i>Anthemis arvensis</i>	-0.16	0.006
<i>Galium aparine</i>	0.16	0.016	<i>Mysotis arvensis</i>	-0.08	0.005	<i>Misopates orontium</i>	-0.31	0.023	<i>Polygonum aviculare</i>	-0.15	0.022
<i>Campanula rapunculoides</i>	0.25	0.011	<i>Polygonum aviculare</i>	0.080.006		<i>Sinapis arvensis</i>	-0.28	0.022	<i>Arrhizum patula</i>	-0.15	0.005
			<i>Stene noctiflora</i>	0.180.008		<i>Medicago lupulina</i>	-0.28	0.016	<i>Campanula rapunculoides</i>	-0.14	0.004

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App. 1, cont.

Altitude Species	Ax 1 score	Fit	Season Species	Ax 1 score	Fit	Year Species	Ax 1 score	Fit	Crop Species	Ax 1 score	Fit
<i>Rumex crispus</i>	0.28	0.023	<i>Spergula arvensis</i>	0.18	0.005	<i>Lathyrus tuberosus</i>	-0.24	0.008	<i>Tripleurospermum inodorum</i>	-0.13	0.012
<i>Poa annua</i>	0.32	0.030	<i>Avena fatua</i>	0.19	0.008	<i>Avena fatua</i>	-0.23	0.011	<i>Viola arvensis</i>	-0.13	0.022
<i>Myosotis arvensis</i>	0.33	0.089	<i>Sonchus arvensis</i>	0.19	0.012	<i>Aethusa cynapium</i>	-0.22	0.010	<i>Setaria viridis</i>	-0.13	0.022
<i>Agerostis soloncheta</i>	0.33	0.012	<i>Equisetum arvense</i>	0.20	0.009	<i>Viola hirsuta</i>	-0.22	0.014	<i>Avena fatua</i>	-0.12	0.003
<i>Viola hirsuta</i>	0.35	0.036	<i>Chenopodium album</i>	0.21	0.031	<i>Galeopsis tetralix</i>	-0.15	0.009	<i>Fallopia convolvulus</i>	-0.07	0.007
<i>Chenopodium uliginosum</i>	0.36	0.016	<i>Convolvulus arvensis</i>	0.24	0.032	<i>Angallis arvensis</i>	-0.10	0.007	<i>Taraxacum sect. Ruderalia</i>	0.11	0.007
<i>Setaria annua</i>	0.38	0.032	<i>Daucus carota</i>	0.26	0.011	<i>Myosotis arvensis</i>	0.11	0.009	<i>Capsella bursa-pastoris</i>	0.12	0.013
<i>Persicaria maculosa</i>	0.38	0.032	<i>Persicaria lapathifolia</i>	0.27	0.020	<i>Stellaria media</i>	0.13	0.013	<i>Lamium purpureum</i>	0.12	0.005
<i>Veronica arvensis</i>	0.39	0.051	<i>Persicaria hydropiper</i>	0.28	0.006	<i>Tripleurospermum inodorum</i>	0.13	0.012	<i>Persicaria maculosa</i>	0.14	0.004
<i>Raphanus raphanistrum</i>	0.41	0.052	<i>Mentha arvensis</i>	0.28	0.021	<i>Veronica persica</i>	0.17	0.021	<i>Raphanus raphanistrum</i>	0.14	0.006
<i>Aethusa arvensis</i>	0.42	0.038	<i>Atriplex panula</i>	0.31	0.023	<i>Euphorbia helioscopia</i>	0.17	0.011	<i>Euphorbia helioscopia</i>	0.19	0.014
<i>Viola sativa</i>	0.42	0.063	<i>Aethusa cynapium</i>	0.31	0.020	<i>Veronica arvensis</i>	0.18	0.011	<i>Stellaria media</i>	0.20	0.029
<i>Stachys palustris</i>	0.46	0.042	<i>Persicaria maculosa</i>	0.33	0.024	<i>Persicaria lapathifolia</i>	0.28	0.022	<i>Geranium pusillum</i>	0.21	0.008
<i>Marricaria discoides</i>	0.51	0.031	<i>Erysimum cheiranthoides</i>	0.33	0.009	<i>Taraxacum sect. Ruderalia</i>	0.28	0.045	<i>Chenopodium polyspermum</i>	0.22	0.005
<i>Achillea millefolium</i>	0.51	0.080	<i>Plantago lanceolata</i>	0.34	0.012	<i>Gallium aparine</i>	0.29	0.051	<i>Lamium amplexicaule</i>	0.22	0.012
<i>Lapsana communis</i>	0.52	0.116	<i>Stachys palustris</i>	0.35	0.023	<i>Echinochloa crus-galli</i>	0.30	0.007	<i>Erodium cicutarium</i>	0.25	0.009
<i>Ranunculus repens</i>	0.53	0.074	<i>Oxalis fontana</i>	0.35	0.010	<i>Lapsana communis</i>	0.32	0.044	<i>Chenopodium album</i>	0.27	0.049
<i>Poa trivialis</i>	0.53	0.025	<i>Anagallis arvensis</i>	0.36	0.086	<i>Sonchus oleraceus</i>	0.33	0.019	<i>Persicaria lapathifolia</i>	0.29	0.023
<i>Potentilla anserina</i>	0.54	0.024	<i>Serratula arvensis</i>	0.38	0.024	<i>Erysimum cheiranthoides</i>	0.34	0.010	<i>Fumaria officinalis</i>	0.32	0.013
<i>Cerastium holosteoides</i>	0.54	0.039	<i>Plantago major</i>	0.38	0.064	<i>Marricaria discoides</i>	0.34	0.014	<i>Lolium perenne</i>	0.42	0.009
<i>Mentha arvensis</i>	0.55	0.075	<i>Medicago lupulina</i>	0.40	0.021	<i>Ranunculus repens</i>	0.34	0.031	<i>Echinochloa crus-galli</i>	0.51	0.021
<i>Persicaria hydropiper</i>	0.64	0.033	<i>Gnaphalium uliginosum</i>	0.41	0.021	<i>Rumex obtusifolius</i>	0.37	0.013	<i>Chenopodium ficifolium</i>	0.51	0.005
<i>Tussilago farfara</i>	0.71	0.053	<i>Microrrhinum minus</i>	0.42	0.010	<i>Chenopodium polyspermum</i>	0.40	0.019	<i>Chenopodium hybridum</i>	0.52	0.012
<i>Galeopsis pubescens</i>	0.72	0.011	<i>Sonchus oleraceus</i>	0.48	0.040	<i>Amaranthus retroflexus</i>	0.41	0.011	<i>Melba neglecta</i>	0.54	0.005
<i>Viola arvensis</i>	0.76	0.036	<i>Trifolium hybridum</i>	0.49	0.006	<i>Lamium purpureum</i>	0.43	0.058	<i>Amaranthus retroflexus</i>	0.60	0.023
<i>Spergula arvensis</i>	0.79	0.094	<i>Galinisoga quadrifida</i>	0.49	0.014	<i>Lactuca hederacea</i>	0.47	0.011	<i>Euphorbia pephus</i>	0.62	0.006
<i>Rumex acetosella</i>	0.81	0.063	<i>Echinochloa crus-galli</i>	0.50	0.021	<i>Agrostis soloncheta</i>	0.51	0.028	<i>Galinisoga parviflora</i>	0.70	0.030
<i>Prunella vulgaris</i>	0.81	0.016	<i>Setaria viridis</i>	0.51	0.014	<i>Geranium dissectum</i>	0.53	0.033	<i>Solanum nigrum</i>	0.71	0.016
<i>Viola septima</i>	0.81	0.010	<i>Chenopodium polyspermum</i>	0.52	0.031	<i>Lactuca serriola</i>	0.55	0.018	<i>Holcicum umbellatum</i>	0.71	0.004
<i>Veronica serpyllifolia</i>	0.83	0.013	<i>Sonchus asper</i>	0.53	0.090	<i>Artemisia vulgaris</i>	0.56	0.028	<i>Bromus sterilis</i>	0.72	0.004
<i>Stellaria graminea</i>	0.88	0.043	<i>Euphorbia exigua</i>	0.54	0.046	<i>Poa trivialis</i>	0.65	0.037	<i>Galinisoga quadrifida</i>	0.74	0.031
<i>Galopha terrebis</i>	0.91	0.326	<i>Anagallis foemina</i>	0.57	0.017	<i>Arctium tomentosum</i>	0.69	0.009	<i>Mercurialis annua</i>	0.80	0.012
<i>Veronica chamaedrys</i>	0.94	0.015	<i>Stachys annua</i>	0.58	0.008	<i>Matva neglecta</i>	0.69	0.008	<i>Amaranthus viridis</i>	0.81	0.003
<i>Phleum pratense</i>	0.96	0.031	<i>Gypsophila muralis</i>	0.58	0.008	<i>Leucanthemum vulgare</i>	0.71	0.015	<i>Armoeracia rusticana</i>	0.83	0.005
<i>Agrostis gigantea</i>	1.04	0.020	<i>Ruscus laevis</i>	0.61	0.007	<i>Rubus caesius</i>	0.71	0.011	<i>Atriplex sagittata</i>	0.87	0.006
<i>Hydroleptium maximum</i>	1.06	0.026	<i>Galinisoga parviflora</i>	0.62	0.024	<i>Poa pratensis</i>	0.79	0.023	<i>Senecio vulgaris</i>	0.89	0.035
<i>Leucanthemum vulgare</i>	1.06	0.033	<i>Alfuga chamaecephys</i>	0.67	0.007	<i>Pastinaca sativa</i>	0.83	0.011	<i>Hibiscus trionum</i>	0.95	0.003
<i>Rumex acetosa</i>	1.20	0.047	<i>Amaranthus retroflexus</i>	0.68	0.030	<i>Euphorbia pephus</i>	0.84	0.012	<i>Atriplex oblongifolia</i>	0.97	0.004
<i>Trifolium hybridum</i>	1.23	0.039	<i>Kickxia spurta</i>	0.71	0.017	<i>Crepis biennis</i>	0.85	0.012	<i>Chenopodium glaucum</i>	0.97	0.010
<i>Rhinanthus elecrolophus</i>	1.28	0.023	<i>Euphorbia falcata</i>	0.74	0.016	<i>Symphitum officinale</i>	0.91	0.024	<i>Urtica urens</i>	1.27	0.008
<i>Lathyrus pratensis</i>	1.32	0.028	<i>Misopates orontium</i>	0.75	0.015	<i>Arrhenatherum elatius</i>	0.95	0.011	<i>Erucastrum gallicum</i>	1.30	0.003
<i>Holcus mollis</i>	1.32	0.087	<i>Solanum nigrum</i>	0.80	0.021	<i>Aegopodium podagraria</i>	0.96	0.031	<i>Eragrostis minor</i>	1.30	0.004
<i>Sanguisorba officinalis</i>	1.38	0.011	<i>Amaranthus hybridus</i>	0.84	0.007	<i>Leonodon autumnalis</i>	1.04	0.009	<i>Aristolochia clematitis</i>	1.36	0.004
<i>Hypericum maculatum</i>	1.43	0.026	<i>Kickxia elatine</i>	0.86	0.024	<i>Lolium perenne</i>	1.06	0.062	<i>Amaranthus blitum</i>	1.60	0.004
<i>Festuca pratensis</i>	1.48	0.037	<i>Amaranthus powellii</i>	0.86	0.010	<i>Urtica dioica</i>	1.12	0.026	<i>Pulegium vulgare</i>	1.78	0.004
<i>Agrostis capillaris</i>	1.63	0.033	<i>Setaria pumila</i>	0.87	0.047	<i>Galinisoga quadrifida</i>	1.14	0.073	<i>Coronopus squamatus</i>	1.81	0.004
<i>Lolium temulentum</i>	2.03	0.010	<i>Salsola verticillata</i>	0.88	0.007	<i>Trisetum flavescens</i>	1.19	0.008	<i>Muscari comosum</i>	1.88	0.003
<i>Equisetum sylvaticum</i>	2.07	0.022	<i>Galeopsis angustifolia</i>	1.00	0.006	<i>Chaerophyllum aromanticum</i>	1.20	0.012	<i>Gegea villosa</i>	2.19	0.006
<i>Achenilla vulgaris</i> s. lat.	2.12	0.063	<i>Carduus nutans</i>	1.71	0.007	<i>Trifolium medium</i>	1.25	0.010	<i>Orrhizogalum boucheanum</i>	2.20	0.003
<i>Melampyrum pratense</i>	3.24	0.018	<i>Achillea setacea</i>	1.97	0.006	<i>Persicaria mitis</i>	1.30	0.015	<i>Tragopogon dubius</i>	2.30	0.007
<i>Rumex arifolius</i>	3.25	0.011	<i>Scabiosa ochroleuca</i>	2.03	0.007	<i>Cerastium glomeratum</i>	1.37	0.020	<i>Cerastium brachyepetalum</i>	2.43	0.007

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